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Should the genomic DNA downstream of the promoter contain, in an antisense orientation, at least a portion of an mRNA or a non-translated RNA encoding a gene product involved in proliferation, then induction of transcription from the promoter will result in detectable inhibition of proliferation.

In the case of *Staphylococcus aureus*, a shotgun library of *Staphylococcus aureus* genomic fragments was cloned into the vector pXyIT5-P15a, which harbors the XyIT5 inducible promoter. The vector was linearized at a unique *Bam*HI site immediately downstream of the XyIT5 promoter/operator. The linearized vector was treated with shrimp alkaline phosphatase to prevent reclosure of the linearized ends. Genomic DNA isolated from *Staphylococcus aureus* strain RN450 was fully digested with the restriction enzyme *Sau*3A, or, alternatively, partially digested with DNase I and "blunt-ended" by incubating with T4 DNA polymerase. Random genomic fragments between 200 and 800 base pairs in length were selected by gel purification. The size-selected genomic fragments were added to the linearized and dephosphorylated vector at a molar ratio of 0.1 to 1, and ligated to form a shotgun library.

The ligated products were transformed into electrocompetent E. coli strain XL1-Blue MRF (Stratagene) and plated on LB medium with supplemented with carbenicillin at 100 μ g/ml. Resulting colonies numbering 5 x 10⁵ or greater were scraped and combined, and were then subjected to plasmid purification.

The purified library was then transformed into electrocompetent *Staphylococcus aureus* RN4220. Resulting transformants were plated on agar containing LB + 0.2% glucose (LBG medium) + chloramphenicol at 15 µg/ml (LBG+CM15 medium) in order to generate 100 to 150 platings at 500 colonies per plating. The colonies were subjected to robotic picking and arrayed into wells of 384 well culture dishes. Each well contained 100µl of LBG + CM15 liquid medium. Inoculated 384 well dishes were incubated 16 hours at 37°C, and each well was robotically gridded onto solid LBG + CM15 medium with or without 2% xylose. Gridded plates were incubated 16 hours at 37°C, and then manually scored for arrayed colonies that were growth-compromised in the presence of xylose.

Arrayed colonies that were growth-sensitive on medium containing 2% xylose, yet were able to grow on similar medium lacking xylose, were subjected to further growth sensitivity analysis as follows: Colonies from the plate lacking xylose were manually picked and inoculated into individual wells of a 96 well culture dish containing LBG + CM15, and were incubated for 16 hours at 37°C. These cultures were robotically diluted 1/100 into fresh medium and allowed to incubate for 4 hours at 37°C, after which they were subjected to serial dilutions in a 384 well array and then gridded onto media containing 2% xylose or media lacking xylose. After growth for 16 hours at 37°C, the arrays that resulted on the two media were compared to each other. Clones that grew similarly at all dilutions on both media were scored as a negative and were no longer considered. Clones that grew on xylose medium but failed to grow at the same serial dilution on the non-xylose plate were given a score based on the differential, i.e. should the clone grow at a

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serial dilution of 10⁴ or less on the xylose plate and grow at a serial dilution of 10⁸ or less on the non-xylose plate, then the corresponding clone received a score of "4" representing the log difference in growth observed.

For Salmonella typhimurium and Klebsiella pneumoniae growth curves were carried out by back diluting cultures 1:200 into fresh media containing 1 mM IPTG or media lacking IPTG and measuring the OD_{450} every 30 minutes (min). To study the effects of transcriptional induction on solid medium, 10^2 , 10^3 , 10^4 , 10^5 , 10^6 , 10^7 and 10^8 fold dilutions of overnight cultures were prepared. Aliquots of from 0.5 to 3 μ l of these dilutions were spotted on selective agar plates with or without 1 mM IPTG. After overnight incubation, the plates were compared to assess the sensitivity of the clones to IPTG.

Nucleic acids involved in proliferation of *Pseudomonas aeruginosa* were identified as follows. Randomly generated fragments of *Pseudomonas aeruginosa* genomic DNA were transcribed from a two-component inducible promoter system. Integrated on the chromosome was the T7 RNA polymerase gene regulated by *lac*UV5/ *lac*O (Brunschwig, E. and Darzins, A. 1992. Gene 111:35-41). On an expression plasmid there was a T7 gene 10 promoter, which is transcribed by T7 RNA polymerase, fused with a *lac*O operator followed by a multiple cloning site. Transcription from this hybrid promoter is inducible by IPTG. Should the genomic DNA downstream of the promoter contain, in an antisense orientation, at least a portion of an mRNA encoding a gene product involved in proliferation, then induction of expression from the promoter will result in detectable inhibition of proliferation.

A shotgun library of *Pseudomonas aeruginosa* genomic fragments was cloned into the vectors pEP5, pEP5S, or other similarly constructed vectors which harbor the T7*lac*O inducible promoter. The vector was linearized at a unique *Sma*I site immediately downstream of the T7*lac*O promoter/operator. The linearized vector was treated with shrimp alkaline phosphatase to prevent reclosure of the linearized ends. Genomic DNA isolated from *Pseudomonas aeruginosa* strain PAO1 was partially digested with DNase I and "blunt-ended" by incubating with T4 DNA polymerase. Random genomic fragments between 200 and 800 base pairs in length were selected by gel purification. The size-selected genomic fragments were added to the linearized and dephosphorylated vector at a molar ratio of 2 to 1, and ligated to form a shotgun library.

The ligated products were transformed into electrocompetent E. coli strain XL1-Blue MRF (Stratagene) and plated on LB medium with carbenicillin at 100 µg/ml or Streptomycin 100 µg/ml. Resulting colonies numbering 5 x 10^5 or greater were scraped and combined, and were then subjected to plasmid purification.

The purified library was then transformed into electrocompetent *Pseudomonas aeruginosa* strain PAO1. Resulting transformants were plated on LB agar with carbenicillin at $100~\mu g/ml$ or Streptomycin 40 $\mu g/ml$ in order to generate 100 to 150 platings at 500 colonies per plating. The colonies were subjected to robotic picking and arrayed into wells of 384 well culture dishes. Each well contained 100 μ l of LB + CB 100 or Streptomycin 40 liquid medium. Inoculated 384 well

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dishes were incubated 16 hours at room temperature, and each well was robotically gridded onto solid LB + CB100 or Streptomycin 40 medium with or without 1 mM IPTG. Gridded plates were incubated 16 hours at 37°C, and then manually scored for arrayed colonies that were growth-compromised in the presence of IPTG.

Arrayed colonies that were growth-sensitive on medium containing 1 mM IPTG, yet were able to grow on similar medium lacking IPTG, were subjected to further growth sensitivity analysis as follows: Colonies from the plate lacking IPTG were manually picked and inoculated into individual wells of a 96 well culture dish containing LB + CB100 or Streptomycin 40, and were incubated for 16 hours at 30°C. These cultures were robotically diluted 1/100 into fresh medium and allowed to incubate for 4 hours at 37°C, after which they were subjected to serial dilutions in a 384 well array and then gridded onto media with and without 1 mM IPTG. After growth for 16 hours at 37°C, the arrays of serially diluted spots that resulted were compared between the two media. Clones that grew similarly at all dilutions on both media were scored as a negative and were no longer considered. Clones that grew on IPTG medium but failed to grow at the same serial dilution on the non-IPTG plate were given a score based on the differential, i.e. should the clone grow at a serial dilution of 10⁴ or less on the IPTG plate and grow at a serial dilution of 10⁸ or less on the IPTG plate, then the corresponding clone received a score of "4" representing the log difference in growth observed.

Following the identification of those vectors that, upon induction, negatively impacted *Pseudomonas aeruginosa* growth or proliferation, the inserts or nucleic acid fragments contained in those vectors were isolated for subsequent characterization. Vectors of interest were subjected to nucleic acid sequence determination.

Nucleic acids involved in proliferation of *E. faecalis* were identified as follows. Randomly generated fragments of genomic DNA were expressed from the vectors pEPEF3 or pEPEF14, which contain the CP25 or P59 promoter, respectively, regulated by the xyl operator/repressor. These plasmids as well as other vectors useful for the expression of nucleic acids in *Enterococcus faecalis* and other Gram positive organisms are described in U.S. Patent Application Serial Number 10/032,393, filed December 21, 2001, the disclosure or which is incorporated herein by reference in its entirety. Should the genomic DNA downstream of the promoter contain, in an antisense orientation, at least a portion of a mRNA encoding a gene product involved in proliferation, then induction of expression from the promoter will result in detectable inhibition of proliferation.

A shotgun library of *E. faecalis* genomic fragments was cloned into the vector pEPEF3 or pEPEF14, which harbor xylose inducible promoters. The vector was linearized at a unique *SmaI* site immediately downstream of the promoter/operator. The linearized vector was treated with alkaline phosphatase to prevent reclosure of the linearized ends. Genomic DNA isolated from *E. faecalis* strain OG1RF was partially digested with DNase I and "blunt-ended" by incubating with T4 DNA polymerase. Random genomic fragments between 200 and 800 base pairs in length were

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selected by gel purification. The size-selected genomic fragments were added to the linearized and dephosphorylated vector at a molar ratio of 2 to 1, and ligated to form a shotgun library.

The ligated products were transformed into electrocompetent E. coli strain TOP10 cells (Invitrogen) and plated on LB medium with erythromycin (Erm) at 150 μ g/ml. Resulting colonies numbering 5 x 10^5 or greater were scraped and combined, and were then subjected to plasmid purification.

The purified library was then transformed into electrocompetent *E. faecalis* strain OG1RF. Resulting transformants were plated on Todd-Hewitt (TH) agar with erythromycin at 10 μg/ml in order to generate 100 to 150 platings at 500 colonies per plating. The colonies were subjected to robotic picking and arrayed into wells of 384 well culture dishes. Each well contained 100 μl of THB + Erm 10 μg/ml. Inoculated 384 well dishes were incubated 16 hours at room temperature, and each well was robotically gridded onto solid TH agar + Erm with or without 5% xylose. Gridded plates were incubated 16 hours at 37°C, and then manually scored for arrayed colonies that were growth-compromised in the presence of xylose.

Arrayed colonies that were growth-sensitive on medium containing 5% xylose, yet were able to grow on similar medium lacking xylose, were subjected to further growth sensitivity analysis. Colonies from the plate lacking xylose were manually picked and inoculated into individual wells of a 96 well culture dish containing THB + Erm 10, and were incubated for 16 hours at 30°C. These cultures were robotically diluted 1/100 into fresh medium and allowed to incubate for 4 hours at 37°C, after which they were subjected to serial dilution on plates containing 5% xylose or plates lacking xylose. After growth for 16 hours at 37°C, the arrays of serially diluted spots that resulted were compared between the two media. Colonies that grew similarly on both media were scored as a negative and corresponding colonies were no longer considered. Colonies on xylose medium that failed to grow to the same serial dilution compared to those on the non-xylose plate were given a score based on the differential. For example, colonies on xylose medium that only grow to a serial dilution of -4 while they were able to grow to -8 on the non-xylose plate, then the corresponding transformant colony received a score of "4" representing the log difference in growth observed.

Following the identification of those vectors that, upon induction, negatively impacted E. faecalis growth or proliferation, the inserts or nucleic acid fragments contained in those expression vectors were isolated for subsequent characterization. The inserts in the vectors of interest were subjected to nucleotide sequence determination.

It will be appreciated that other restriction enzymes and other endonucleases or methodologies may be used to generate random genomic fragments. In addition, random genomic fragments may be generated by mechanical shearing. Sonication and nebulization are two such techniques commonly used for mechanical shearing of DNA.

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EXAMPLE 2

Nucleotide Sequence Determination of Identified Clones Transribing Nucleic Acid Fragments with

Detrimental Effects on Escherichia coli, Staphylococcus aureus, Enterococcus faecalis, Klebsiella

pneumoniae, Pseudomonas aeruginosa or Salmonella typhimurium Proliferation

Plasmids from clones that received a dilution plating score of "2" or greater were isolated to obtain the genomic DNA insert responsible for growth inhibition as follows.

The nucleotide sequences of the nucleic acid sequences which inhibited the growth of *Escherichia coli* were determined using plasmid DNA isolated using QIAPREP (Qiagen, Valencia, CA) and methods supplied by the manufacturer. The primers used for sequencing the inserts were 5'-TGTTTATCAGACCGCTT - 3' (SEQ ID NO: 78586) and 5'-ACAATTTCACACAGCCTC - 3' (SEQ ID NO: 78587). These sequences flank the polylinker in pLEX5BA.

The nucleotide sequences of the nucleic acid sequences which inhibited the growth of *Staphylococcus aureus* were determined as follows. *Staphylococcus aureus* were grown in standard laboratory media (LB or TB with 15 ug/ml Chloramphenicol to select for the plasmid). Growth was carried out at 37°C overnight in culture tubes or 2 ml deep well microtiter plates.

Lysis of *Staphylococcus aureus* was performed as follows. Cultures (2-5 ml) were centrifuged and the cell pellets resuspended in 1.5 mg/ml solution of lysostaphin (20 μ l/ml of original culture) followed by addition of 250 μ l of resuspension buffer (Qiagen). Alternatively, cell pellets were resuspended directly in 250 μ l of resuspension buffer (Qiagen) to which 5-20 μ l of a 1 mg/ml lysostaphin solution were added.

DNA was isolated using Qiagen miniprep kits or Wizard (Qiagen) miniprep kits according to the instructions provided by the manufacturer.

The genomic DNA inserts were amplified from the purified plasmids by PCR as follows.

1 μl of Qiagen purified plasmid was put into a total reaction volume of 25 μl Qiagen Hot Start PCR mix. For *Staphylococcus aureus*, the following primers were used in the PCR reaction: pXylT5F: CAGCAGTCTGAGTTATAAAATAG (SEQ ID NO: 78588)

LexL TGTTTTATCAGACCGCTT (SEQ ID NO: 78589)

Similar methods were conducted for *Salmonella typhimurium* and *Klebsiella pneumoniae*. For *Salmonella typhimurium* and *Klebsiella pneumoniae* the following primers were used:

30 5' - TGTTTTATCAGACCGCTT - 3' (SEQ ID NO: 78589) and

5'-ACAATTTCACACAGCCTC-3' (SEQ ID NO: 78587)

PCR was carried out in a PE GenAmp with the following cycle times:

Step 1. 95° C 15 min

Step 2. 94° C 45 sec

35 Step 3. 54° C 45 sec

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Step 4. 72° C 1 minute

Step 5. Return to step 2, 29 times

Step 6. 72° C 10 minutes

Step 7. 4° C hold

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The PCR products were cleaned using Qiagen Qiaquick PCR plates according to the manufacturer's instructions.

For *Pseudomonas aeruginosa*, plasmids from transformant colonies that received a dilution plating score of "2" or greater were isolated to obtain the genomic DNA insert responsible for growth inhibition as follows. *Pseudomonas aeruginosa* were grown in standard laboratory media (LB with carbenicillin at 100 μg/ml or Streptomycin 40 μg/ml to select for the plasmid). Growth was carried out at 30°C overnight in 100 ul culture wells in microtiter plates. To amplify insert DNA 2 ul of culture were placed into 25 ul Qiagen Hot Start PCR mix. PCR reactions were in 96 well microtiter plates. For plasmid pEP5S the following primers were used in the PCR reaction:

T7L1+: GTCGGCGATATAGGCGCCAGCAACCG (SEQ ID NO: 78590)

pStrA3: ATAATCGAGCATGAGTATCATACG (SEQ ID NO: 78591)

PCR was carried out in a PE GenAmp with the following cycle times:

Step 1. 95° C 15 min

15 Step 2. 94° C 45 sec

Step 3. 54° C 45 sec

Step 4. 72° C 1 minute

Step 5. Return to step 2, 29 times

Step 6. 72° C 10 minutes

20 Step 7. 4° C hold

The PCR products were cleaned using Qiagen Qiaquick PCR plates according to the manufacturer's instructions.

The purified PCR products were then directly cycle sequenced with Qiagen Hot Start PCR mix. The following primers were used in the sequencing reaction:

25 T7/L2: ATGCGTCCGGCGTAGAGGAT (SEQ ID NO: 78592)

PCR was carried out in a PE GenAmp with the following cycle times:

Step 1. 94° C 15 min

Step 2. 96° C 10 sec

Step 3. 50° C 5 sec

30 Step 4. 60 C 4 min

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Step 5. Return to step 2, 24 times

Step 6. 4° C hold

The PCR products were cleaned using Qiagen Qiaquick PCR plates according to the manufacturer's instructions.

For *E. faecalis*, plasmids from transformant colonies that received a dilution plating score of "2" or greater were isolated to obtain the genomic DNA insert responsible for growth inhibition as follows. *E. faecalis* were grown in THB 10 µg/ml Erm at 30°C overnight in 100 ul culture wells

in microtiter plates. To amplify insert DNA 2 ul of culture were placed into 25 μ l Qiagen Hot Start PCR mix. PCR reactions were in 96 well microtiter plates. The following primers were used in the PCR reaction:

pXyIT5: CAGCAGTCTGAGTTATAAAATAG (SEQ ID NO: 78588) and the

5 pEP/pAK1 primer.

PCR was carried out in a PE GenAmp with the following cycle times:

Step 1. 95° C 15 min

Step 2. 94° C 45 sec

Step 3. 54° C 45 sec

10 Step 4. 72° C 1 minute

Step 5. Return to step 2, 29 times

Step 6. 72° C 10 minutes

Step 7. 4° C hold

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The PCR products were cleaned using Qiagen Qiaquick PCR plates according to the manufacturer's instructions.

The purified PCR products were then directly cycle sequenced with Qiagen Hot Start PCR mix. The following primers were used in the PCR reaction:

pXyIT5: CAGCAGTCTGAGTTATAAAATAG (SEQ ID NO: 78588)

PCR was carried out in a PE GenAmp with the following cycle times:

20 Step 1. 94° C 15 min

Step 2. 96° C 10 sec

Step 3. 50° C 5 sec

Step 4. 60° C 4 min

Step 5. Return to step 2, 24 times

25 Step 6. 4° C hold

The PCR products were cleaned using Qiagen Qiaquick PCR plates according to the manufacturer's instructions.

The amplified genomic DNA inserts from each of the above procedures were subjected to automated sequencing. Sequence identification numbers (SEQ ID NOs) and clone names for the identified inserts are listed in Table IA and discussed below.

TABLE IA

Clone Name	S1M10000025G06	S1M10000025H06	S1M10000025H07	S1M10000025A08	S1M10000025D08	S1M10000025F08	S1M10000025H08	S1M10000025A09	S1M10000025B09	S1M10000025C09	S1M10000025D09	SIM10000025E09	S1M10000025F09	S1M10000025A10	S1M10000025C10	S1M10000025D10	S1M10000025F10	SIM10000025G10	S1M10000025H10	S1M10000025C11	S1M10000025E11	S1M10000025B12	S1M10000025F12	S1M10000026C01	S1M10000026E01	S1M10000026F01	S1M10000026G01	S1M10000026H01	S1M10000026A02	S1M10000026B02	S1M10000026H02	S11M10000020B03	CONTRACTOR
SeqID	4969	4970	4971	4972	4973	4974	4975	4976	4977	4978	4979	4980	4981	4982	4983	4984	4985	4986	4987	4988	4989	4990	4991	4992	4993	4994	4995	4996	4997	4998	4999	2000	2001
Clone Name	P1M10000105C04	P1M10000105D04	P1M10000105C05	P1M10000105B06	P1M10000105C08	P1M10000105H08	P1M10000105D09	P1M10000110E01	P1M10000110F01	P1M10000110G01	P1M10000110B02	P1M10000110B03	P1M10000110F03	P1M10000110G03	P1M10000110D04	P1M10000110F04	P1M10000110B05	P1M10000110E05	P1M10000110B07	P1M10000110B08	P1M10000110F08	P1M10000110A09	P1M10000110E09	P1M10000110F09	P1M10000100F01	P1M10000098A02	P1M10000098B02	P1M10000098A03	P1M10000098D03	P1M10000098E04	P1M10000098G04	P1M10000098A05	P1M10000098C05
SeqID	3727	3728	3729	3730	3731	3732	3733	3734	3735	3736	3737	3738	3739	3740	3741	3742	3743	3744	3745	3746	3747	3748	3749	3750	3751	3752	3753	3754	3755	3756	3757	3758	3759
Clone Name	E1M10000260G02	E1M10000260F04	E1M10000260A05	E1M10000260C05	E1M10000260E05	E1M10000260C07	E1M10000260G07	E1M10000260B08	E1M10000260D08	E1M10000260E08	E1M10000260E09	E1M10000260C10	E1M10000260D10	E1M10000260E10	E1M10000260G10	E1M10000260H10	E1M10000260H11	E1M10000260B12	E1M10000260D12	E1M10000260G12	E1M10000261F01	E1M10000261B02	E1M10000261H02	E1M10000261G04	E1M10000261H05	E1M10000261G06	E1M10000261H06	E1M10000261D08	E1M10000261F08	E1M10000261C09	E1M10000261H09	E1M10000261E10	E1M10000262E01
SeqID	2485	2486	2487	2488	2489	2490	2491	2492	2493	2494	2495	2496	2497	2498	2499	2500	2501	2502	2503	2504	2505	2506	2507	2508	2509	2510	2511	2512	2513	2514	2515	2516	2517
Clone Name	P33-1.C22	X3S107-17	P35-7	X3S118-9	X3S163-1	X3S204-7	X3S177-4	P342-3	SC21.1	SC17.1	SC13.1	MC9.6	Z60-P16	Z86-I21	E1M10000109A02	E1M10000109A11	E1M10000101F05	E1M10000101D06	E1M10000101A07	E1M10000101H07	E1M10000101H09	E1M10000101C12	E1M10000103B04	E1M10000103D11	E1M10000110G01	E1M10000110H01	E1M10000110E09	E1M10000110A12	E1M10000112F05	E1M10000113F02	E1M10000113A11	E1M10000111C03	E1M10000111E04
SeqID	1243	1244	1245	1246	1247	1248	1249	1250	1251	1252	1253	1254	1255	1256	1257	1258	1259	1260	1261	1262	1263	1264	1265	1266	1267	1268	1269	1270	1271	1272	1273	1274	1275
Clone Name	E3M10000001B01	E3M10000001A02	E3M10000001B02	E3M10000001C02	E3M10000001D02	E3M10000001E02	E3M10000001F02	E3M10000001G02	E3M10000001H02	E3M10000001E03	E3M10000001G03	E3M10000001H03	E3M10000001D04	E3M10000001E04	E3M10000001F04	E3M10000001G04	E3M10000001H04	E3M10000001B05	E3M10000001D05	E3M10000001G05	E3M10000001A06	E3M10000001F06	E3M10000001B08	E3M10000001E08	E3M10000001C09	E3M10000001D09	E3M10000001E09	E3M10000001B10	E3M10000004D01	E3M10000004G01	E3M10000004D02	E3M10000004C03	E3M10000004A04
SeaID		2	3	4	5	9		- ∞	6	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33

Clone Name SIM10000026G03 SIM10000026H03 SIM10000026H04 SIM10000026CG04 SIM10000026G04 SIM10000026G04 SIM10000026G05 SIM10000026G05 SIM10000026G05 SIM10000026G06 SIM10000026G06 SIM10000026G06 SIM10000026G06 SIM10000026G07	S1M10000026G08 S1M10000026E09 S1M10000026E09 S1M10000026H09 S1M10000026H10
SeqID 5002 5003 5004 5004 5005 5005 5004 5007 5008 5009 5010 5011 5012 5012 5013 5010 5010 5010 5020 5020 5020 5020 5020	5032 5033 5034 5035 5036 5036
Clone Name PIM10000098G06 PIM10000098H06 PIM10000098H06 PIM10000098H09 PIM10000098H09 PIM10000099H11 PIM10000099G03 PIM10000099G12 PIM10000099G10 PIM10000099F10 PIM10000099F10 PIM10000106H03 PIM10000106H03 PIM10000106H03 PIM10000106H03 PIM10000106H03 PIM10000106H03 PIM10000106H03 PIM10000106H03 PIM10000107C03 PIM10000107C03 PIM10000107C04 PIM10000107C09 PIM10000107C09	P1M10000107H10 P1M10000108C01 P1M10000108A02 P1M10000108B02 P1M10000108B03
SeqID 3760 3761 3762 3765 3765 3766 3765 3766 3770 3771 3775 3775 3775 3778 3778 3778 3780 3788 3788 3788 3788	3790 3791 3792 3793 3794 3795
Clone Name E1M10000262C02 E1M10000262E02 E1M10000262E03 E1M10000262C04 E1M10000262C05 E1M10000262C05 E1M10000262E07 E1M10000262E07 E1M10000262E07 E1M10000262E07 E1M10000262E07 E1M10000263E10 E1M10000263E11	E1M10000264F03 E1M10000264A04 E1M10000264B04 E1M10000264C04 E1M10000264E04
SeqID 2518 2519 2520 2521 2521 2522 2523 2524 2524 2528 2528 2528 2529 2539 2539 2539 2534 2534 2534 2534 2534 2534 2534 2534	2548 2549 2550 2551 2552 2553
Clone Name E1M10000111F09 E1M10000115H01 E1M10000115G04 E1M10000115G04 E1M10000116G02 E1M10000106G02 E1M10000106F05 E1M10000106H05 E1M10000106G02 E1M10000106G09 E1M10000106G09 E1M10000106G10 E1M100001023B03 E1M10000123B03 E1M10000123H10 E1M10000123H11 E1M10000123H11 E1M10000123H11 E1M10000123H11 E1M10000107G02 E1M10000107G02 E1M10000107G02 E1M10000107G03 E1M10000107G03	E1M10000107F09 E1M10000107H09 E1M10000117C12 E1M10000118C04 E1M10000118B05 E1M10000118C05
SeqID 1276 1277 1277 1277 1280 1280 1283 1284 1288 1288 1289 1290 1290 1290 1290 1290 1290 1290 129	1306 1307 1308 1309 1310
	E3M10000006G12 E3M10000006G12 E3M10000007F01 E3M10000007A02 E3M10000007A02
SeqID 36 38 39 39 39 39 39 39 39 39 39 39 39 39 39	64 65 67 67 69

Clone Name	S1M10000026B10	SIM10000026D10	S1M10000026E10	S1M10000026F10	S1M10000026G10	S1M10000026H10	S1M10000026A11	S1M10000026B11	S1M10000026C11	S1M10000026E11	S1M10000026B12	S1M10000026C12	SIM10000026D12	SIM10000026E12	S1M10000026F12	S1M10000026G12	S1M10000027G01	S1M10000027C02	S1M10000027D02	S1M10000027E02	S1M1000002/F02	S1M1000002/H02	S1M1000002/D03	S1M1000002/E03	S1M1000002/G03	S1M1000002/A04	S1M1000002/C04	S1M1000002/G04	SIM1000002/H04	SIM1000002/A05	S1M1000002/C05	SIMIO00002/D03	SIM1000002/E05	S1M1000002/F05	S1M1000002/G05	
SeqID	5038	5039	5040	5041	5042	5043	5044	5045	5046	5047	5048	5049	2050	5051	5052	5053	5054	5055	2056	5057	5058	5059	2060	5061	5062	5063	5064	5065	2066	5067	5068	5069	5070	5071	2/05	le/ne
Clone Name	P1M10000108G04	P1M10000108E05	P1M10000108F05	P1M10000108F06	P1M10000108G06	P1M10000109A02	P1M10000109C03	P1M10000109E03	P1M10000109D04	P1M10000109A05	P1M10000109B08	P1M10000109H09	P1M10000109E10	P1M10000109F10	P1M10000109E11	P1M10000109B12	S4M10000001C01	S4M10000002G04	S4M10000002B06	S4M10000002G08	S4M10000002B09	S4M10000019H06	S4M10000008H10	S4M10000009E03	S4M10000009C06	S4M10000009E07	S4M10000009G08	S4M10000009B11	S4M10000009F11	S4M100000009G11	S4M10000010F04	S4M10000010H04	S4M10000010B05	S4M10000010D07	S4M10000010D08	S4M10000010B09
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Clone Name	E1M10000118G06	E1M10000119D02	E1M10000119D03	E1M10000119A04	E1M10000131H01	E1M10000131F04	E1M10000131C06	E1M10000131B07	E1M10000131C07	E1M10000131A10	E1M10000131G10	E1M10000135B02	E1M10000132C01	E1M10000132F02	E1M10000132H04	E1M10000132G08	E1M10000133A06	E1M10000133B08	E1M10000133D09	E1M10000144B01	E1M10000144C02	E1M10000144E03	E1M10000144F03	E1M10000144B06	E1M10000144G06	E1M10000144G07	E1M10000144A08	E1M10000144C10	E1M10000145E01	E1M10000146H01	E1M10000146D02	E1M10000146E05	E1M10000124E02	E1M10000124G03	E1M10000124G04	E1M10000124C05
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Clone Name	E3M10000007B03	E3M10000007C03																			_						E3M10000010G07		E3M10000010G09	E3M10000010G10		E3M10000011B03	E3M10000011D03			E3M10000011B09
SeaID	70	7	77	73	74	75	9/	77	78	0, 0,	`	81	82	83	8 8	85	98	87	× ×	68	06	91	92	93	94	95	96	. 97	98	66	100	101	102	103	104	105

Clone Name	\$1M1000002/B06	90072000011A12	S11M10000027E06	SIMILU000002/E00	S1M1000002/F00	SIM1000002/G00	S1M10000027H06	S1M1000002/B0/	S1M10000027D07	S1M10000027E07	S1M10000027G07	S1M1000002/H0/	S1M110000027A001	S1M1000002/B08	S1M1000002/C08	S1M1000002/D08	SIM1000002/E08	S1M1000002/F08	S1M1000002/G08	SIM1000002/H08	S1M1000002/B09	S1M1000002/C09	S1M1000002/D09	S1M10000027E09	S1M1000002/F09	S1M1000002/G09	S1M1000002/HU9	S1M1000002/D10	SIM1000002/H10	SIM1000002/A11	SIM1000002/B11			SIM1000002/G11		
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Clone Name	S4M10000025E05	S4M10000025H07	S4M10000025A11	S4M10000025F12	S4M10000026C01	S4M10000026E03	S4M10000026D04	S4M10000026B10	S4M10000026E12	S4M10000027E02	S4M10000027C10	S4M10000029B12	S4M10000029D12	S4M10000030F06	S4M10000030F07	S4M10000032F01	S4M10000032G01	S4M10000032F03	S4M10000034A02	S4M10000034C05	S4M10000034H05	S4M10000034A06	S4M10000034A09	S4M10000034H09	S4M10000035B01	S4M10000035D01	S4M10000035F02	S4M10000035E03	S4M10000035B06	S4M10000035A09	S4M10000036F06	S4M10000036B09	S4M10000036H11	S4M10000037A03	S4M10000037A08	S4M10000037H09
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SeaID	1384	1385	1386	1387	1388	1389	1390	1391	1392	1393	1394	1395	1396	1397	1398	1399	1400	1401	1402	1403	1404	1405	1406	1407	1408	1409	1410	1411	1412	1413	1414	1415	1416	1417	1418	1419
Clone Name	E3M1		` '		· 						· 		E3M10000019B06													E3M10000021G11	E3M10000021H11								5 E3M10000022C06	E3M10000022F06
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Clone Name	S1M10000028B08	S1M10000028C08	S1M10000028D08	S1M10000028E08	S1M10000028G08	S1M10000028H08	S1M10000028B09	S1M10000028D09	S1M10000029F01	S1M10000029G01	S1M10000029H01	S1M10000029A02	S1M10000029502	SIM10000029C02	S1M10000029D02	S1M10000029E02	S1M10000029F02	S1M10000029G02	S1M10000029A03	S1M10000029B03	S1M10000029C03	S1M10000029G03	S1M10000029A04	S1M10000029B04	S1M10000029F04	S1M10000029G04	S1M10000029B05	S1M10000029C05	S1M10000029D05	S1M10000029E05	S1M10000029G05	S1M10000029H05	S1M10000029B06	S1M100000029H00	S1M10000029C07	
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SeqID	5182	5183	5184	5185	5186	5187	5188	5189	5190	5191	5192	5193	5194	5195	5196	5197	5198	5199	5200	5201	5202	5203	5204	5205	5206	5207	5208	5209	5210	5211	5212	5213	5214	5215	5210	11770
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Clone Name	S1M10000030E12	S1M10000030G12	S1M10000031B01	S1M10000031H01	S1M10000031B02	S1M10000031E02	S1M10000031F02	S1M10000031G02	S1M10000031H02	S1M10000031A03	S1M10000031E03	S1M10000031F03	S1M10000031G03	S1M10000031A04	S1M10000031B04	S1M10000031C04	S1M10000031E04	S1M10000031F04	S1M10000031G04	S1M10000031F05	S1M10000031D06	S1M10000031G06	S1M10000031H06	S1M10000031C07	S1M10000031D07	S1M10000031E07	S1M10000031A08	S1M10000031D08	S1M10000031E08	S1M10000031F08	S1M10000031C09	S1M10000031D09	S1M10000031G09	S1M10000031H09	S1M10000031A10	S1M10000031C10
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Clone Name	S1M10000024D11	S1M10000024G12	S1M10000025B01	S1M10000025C01	S1M10000025D01	S1M10000025E01	S1M10000025B02	S1M10000025A03	S1M10000025B03	S1M10000025C03	S1M10000025D03	S1M10000025F03	S1M10000025D04	S1M10000025E04	S1M10000025G04	· S1M10000025B05	S1M10000025C05	S1M10000025F05	S1M10000025H05	S1M10000025B06	S1M10000025D06
SeqID	4948	4949	4950	4951	4952	4953	4954	4955	4956	4957	4958	4959	4960	4961	4962	4963	4964	4965	4966	4967	4968
Clone Name	P1M10000094H04	P1M10000094A10	P1M10000095C01	P1M10000095E04	P1M10000095G04	P1M10000095C09	P1M10000102E05	P1M10000102B07	P1M10000103B05	P1M10000103D06	P1M10000103E08	P1M10000104A02	P1M10000104H02	P1M10000104A03	P1M10000104E03	P1M10000104F07	P1M10000104D11	P1M10000105D01	P1M10000105E02	P1M10000105C03	P1M10000105G03
SeqID	3706	3707	3708	3709	3710	3711	3712	3713	3714	3715	3716	3717	3718	3719	3720	3721	3722	3723	3724	3725	3726
Clone Name	E1M10000258G04	E1M10000258C05	E1M10000258D05	E1M10000258F05	E1M10000258G05	E1M10000258A06	E1M10000258D06	E1M10000258B07	E1M10000258G07	E1M10000258G08	E1M10000258B09	E1M10000258D09	E1M10000258F10	E1M10000258C11	E1M10000258F11	E1M10000259C03	E1M10000259B04	E1M10000259E04	E1M10000259E05	E1M10000259B12	E1M10000260E02
SeqID	2464																				2484
Clone Name	P347.2	P31-11-J20	P336-14.F20	P31-27-M1	P338-4.M21	P334-8.L7	. P31-2-E16	P335-3.J14	P334-5.H2	P31-33-N2	P332-11.C20	869.A23	P317-2.A3	P326-9.K2	P323-8.P1	P35-8	P36-13.E2	P38-1.G20	P327-50.M10	P328-8.D21	P328-20.P20
SeqID	1222	1223	1224	1225	1226	1227	1228	1229	1230	1231	1232	1233	1234	1235	1236	1237	1238	1239	1240	1241	1242

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EXAMPLE 3

Comparison Of Isolated Nucleic Acids to Known Sequences

The nucleotide sequences of the subcloned fragments from Escherichia coli, Staphylococcus aureus, Enterococcus faecalis, Klebsiella pneumoniae, Pseudomonas aeruginosa and Salmonella typhimurium obtained from the expression vectors discussed above were compared to known sequences from Escherichia coli, Staphylococcus aureus, Enterococcus faecalis, Klebsiella pneumoniae, Pseudomonas aeruginosa, Salmonella typhimurium and other microorganisms as follows. First, to confirm that each clone originated from one location on the chromosome and was not chimeric, the nucleotide sequences of the selected clones were compared against the Escherichia coli, Staphylococcus aureus, Enterococcus faecalis, Klebsiella pneumoniae, Pseudomonas aeruginosa or Salmonella typhimurium genomic sequences to align the clone to the correct position on the chromosome. The NCBI BLASTN v 2.0.9 program was used for this comparison, and the incomplete Staphylococcus aureus genomic sequences licensed from TIGR, as well as the NCBI nonredundant GenBank database were used as the source of genomic data. Salmonella typhimurium sequences were compared to sequences available from the Genome Sequencing Center (http://genome.wustl.edu/gsc/salmonella.shtml), and the Sanger Centre (http://www.sanger.ac.uk/projects/S typhi). Pseudomonas aeruginosa sequences were compared to a proprietary database and the NCBI GenBank database. The E. faecalis sequences were compared to a proprietary database.

The BLASTN analysis was performed using the default parameters except that the filtering was turned off. No further analysis was performed on inserts which resulted from the ligation of multiple fragments.

In general, antisense molecules and their complementary genes are identified as follows. First, all possible full length open reading frames (ORFs) are extracted from available genomic databases. Such databases include the GenBank nonredundant (nr) database, the unfinished genome database available from TIGR and the PathoSeq database developed by Incyte Genomics. The latter database comprises over 40 annotated bacterial genomes including complete ORF analysis. If databases are incomplete with regard to the bacterial genome of interest, it is not necessary to extract all ORFs in the genome but only to extract the ORFs within the portions of the available genomic sequences which are complementary to the clones of interest. Computer algorithms for identifying ORFs, such as GeneMark, are available and well known to those in the art. Comparison of the clone DNA to the complementary ORF(s) allows determination of whether the clone is a sense or antisense clone. Furthermore, each ORF extracted from the database can be compared to sequences in well annotated databases including the GenBank (nr) protein database, SWISSPROT and the like. A description of the gene or of a closely related gene in a closely related microorganism is often available in these databases. Similar methods are used to identify antisense clones corresponding to genes encoding non-translated RNAs.

In order to generate the gene identification data compiled in Table IB, each of the cloned nucleic acid sequences discussed above corresponding to SEQ ID NO.s 1-6213 was used to identify the corresponding *Escherichia coli, Staphylococcus aureus, Enterococcus faecalis, Klebsiella pneumoniae, Pseudomonas aeruginosa* or *Salmonella typhimurium* ORFs in the PathoSeq v.4.1 (March 2000 release) database of microbial genomic sequences. For this purpose, the NCBI BLASTN 2.0.9 computer algorithm was used. The default parameters were used except that filtering was turned off. The default parameters for the BLASTN and BLASTX analyses were:

Expectation value (e)=10

Alignment view options: pairwise

Filter query sequence (DUST with BLASTN, SEG with others)=T

Cost to open a gap (zero invokes behavior)=0

Cost to extend a gap (zero invokes behavior)=0

X dropoff value for gapped alignment (in bits) (zero invokes behavior)=0

Show GI's in deflines=F

Penalty for a nucleotide mismatch (BLASTN only)=!3

Reward for a nucleotide match (BLASTN only)=1

Number of one-line descriptions (V)=500

Number of alignments to show (B)=250

Threshold for extending hits=default

20 Perform gapped alignment (not available with BLASTX)=T

Query Genetic code to use=1

DB Genetic code (for TBLAST[nx] only=1

Number of processors to use=1

SeqAlign file

25 Believe the query defline=F

Matrix=BLOSUM62

Word Size= default

Effective length of the database (use zero for the real size)=0

Number of best hits from a region to keep=100

30 Length of region used to judge hits=20

Effective length of the search space (use zero for the real size)=0

Query strands to search against database (for BLAST[nx] and TBLASTX), 3 is both, 1 is top, 2 is bottom=3

Produce HTML output=F

Troduce IIIvin output

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Alternatively, ORFs were identified and refined by conducting a survey of the public and private data sources. Full-length gene protein and nucleotide sequences for these organisms were assembled from various sources. For *Pseudomonas aeruginosa*, gene sequences were adopted from the Pseudomonas genome sequencing project (downloaded from http://www.pseudomonas.com). For *Klebsiella pneumoniae*, *Staphylococcus aureus*, *Streptococcus pneumoniae* and *Salmonella typhi*, genomic sequences from PathoSeq v 4.1 (Mar 2000 release) was reanalyzed for ORFs using the gene finding software GeneMark v 2.4a, which was purchased from GenePro Inc. 451 Bishop St., N.W., Suite B, Atlanta, GA, 30318, USA.

Antisense clones were identified as those clones for which transcription from the inducible promoter would result in the expression of an RNA antisense to a complementary ORF, intergenic

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or intragenic sequence. Those clones containing single inserts and that caused growth sensitivity upon induction are listed in Table IA.

The gene descriptions in the PathoSeq database derive from annotations available in the public sequence databases described above. Where a clone was found to share significant sequence identity to two or more adjacent ORFs, it was listed once for each ORF and the PathoSeq information for each ORF was compiled in Table IB.

Table IA lists the SEQ ID NOs. and clone names of the inserts which inhibited proliferation. This information was used to identify the ORFs (SEQ ID NOs.: 6214-42397) whose gene products (SEQ ID NOs. 42398-78581) were inhibited by the nucleic acids comprising the nucleotide sequences of SEQ ID NOs. 1-6213. Table IB lists the clone name and the PathoSeq Locus containing the clone.

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TABLE IB

Clone Name	Gene	Clone Name	Gene	Clone Name	Gene
Cione Ivallie	LocusID	Cione ivanie	LocusID		LocusID
E3M10000001B01	EFA205257	E1M10000233C05	ECO103161	S1M10000005E05	SAU802496
E3M10000001B01	EFA205258	E1M10000233H05	ECO103224	S1M10000005C06	SAU802121
E3M10000001201	EFA205257	E1M10000233H05	ECO103225	S1M10000005D06	SAU801183
E3M10000001A02	EFA205258	E1M10000233D08	ECO103185	S1M10000005D06	SAU801184
E3M10000001102	EFA205225	E1M10000233F08	ECO103265	S1M10000005A07	SAU800967
E3M10000001B02	EFA201977	E1M10000233F08	ECO103266	S1M10000005B07	SAU802496
E3M10000001B02	EFA203137	E1M10000233A09	ECO104092	S1M10000005D07	SAU801264
E3M10000001E02	EFA200840	E1M10000233A09	ECO104093	S1M10000005A08	SAU802496
E3M10000001D02	EFA202003	E1M10000233E09	ECO103238	S1M10000005B08	SAU800548
E3M10000001E02	EFA200840	E1M10000233E09	ECO103239	S1M10000005D08	SAU800607
E3M10000001E02	EFA200807	E1M10000233F09	ECO103886	S1M10000005E08	SAU802496
E3M10000001102	EFA205257	E1M10000233D10	ECO103242	S1M10000005B09	SAU800122
E3M10000001G02	EFA205258	E1M10000233D10	ECO103243	S1M10000005C09	SAU801481
E3M10000001G02	EFA200811	E1M10000233H10	ECO100094	S1M10000005D09	SAU800542
E3M10000001H02	EFA201987	E1M10000234E01	ECO103884	S1M10000005A10	SAU801723
E3M10000001E03	EFA205258	E1M10000234B02	ECO103886	S1M10000005A10	SAU801722
E3M10000001E03	EFA201987	E1M10000234G02	ECO103233	S1M10000005A11	SAU801644
E3M10000001G03	EFA205258	E1M10000234G02	ECO103234	S1M10000005C11	SAU801113
E3M10000001H03	EFA201987	E1M10000234C05	ECO103181	S1M10000005D11	SAU800547
E3M10000001H03	EFA205258	E1M10000234C07	ECO103844	S1M10000005E11	SAU800155
E3M100000011103	EFA201980	E1M10000234C08	ECO103878	S1M10000005B12	SAU802160
E3M10000001D04	EFA201981	E1M10000234C08	ECO204942	S1M10000005B12	SAU603460
E3M10000001D04	EFA205229	E1M10000234E08	EC0204942 EC0103461	S1M10000005D12	SAU801644
E3M10000001D04	EFA203229 EFA201028	E1M10000234H08	ECO103226	S1M10000005E12	SAU801264
E3M10000001E04	EFA201028 EFA200811	E1M10000234F09	ECO103225	S1M10000006B02	SAU800381
E3M10000001F04	EFA200811 EFA201993	E1M10000234F09	ECO103033	S1M10000006E02	SAU802496
	EFA201993 EFA201980	E1M10000234D10	ECO100876	S1M10000006F02	SAU802160
E3M10000001H04 E3M10000001H04	EFA201980 EFA201981	E1M10000234G10	ECO100880	S1M10000006G02	SAU802100
E3M10000001H04	EFA201981 EFA205229	E1M10000234B12 E1M10000235D01	ECO102233	S1M10000006A03	SAU802496
E3M10000001H04	EFA203229 EFA201993	E1M10000235D01	ECO102233	S1M100000007103	SAU802655
E3M10000001B03	EFA201993 EFA201974	E1M10000235A03	ECO100738	S1M10000006D03	SAU801740
E3M10000001D05	EFA201974 EFA201975	E1M10000235H03	ECO103880	S1M10000000E03	SAU801740
E3M10000001D03	EFA201973	E1M10000235E04	ECO103230 ECO103886	S1M10000006E03	SAU801230
E3M10000001G05	EFA202001	E1M10000235F06	ECO103880 ECO103481	S1M10000000103	SAU801275
	EFA202003	E1M10000235B08	ECO103481 ECO103885	S1M10000006A04	SAU801273
E3M10000001A06 E3M10000001F06	EFA201028	E1M10000235E08	ECO103863 ECO103161	S1M10000000A04	SAU802496
l	EFA201028	E1M10000235E08	ECO103101 ECO101848	S1M10000006C04	SAU802158
E3M10000001B08 E3M10000001E08	EFA201028 EFA200807	E1M10000235H09	ECO101848	S1M10000006E04	SAU801089
E3M10000001E08	EFA200807 EFA200839	E1M10000235H09	ECO103481 ECO103482	S1M10000006E04	SAU801644
	EFA200839 EFA201987	E1M10000235H09	ECO103482 ECO100886	S1M10000000104	SAU801044 SAU801740
E3M10000001D09 E3M10000001D09	EFA201987	E1M10000235B10	ECO100880 ECO102299	S1M10000000G04	SAU801740
E3M10000001E09	EFA203238 EFA201987	E1M10000235A11 E1M10000235F12	ECO102233	S1M10000006A05	SAU802223
E3M10000001E09	EFA201987	E1M10000235F12	ECO103233 ECO103234	S1M10000006D05	SAU802496
			ECO103234 ECO100095	S1M10000000B03	SAU801256
E3M10000001B10	EFA205257 EFA205258	E1M10000236E01 E1M10000236A02	ECO100093 ECO102340	S1M10000006G03	
E3M10000001B10 E3M10000004D01	EFA205258 EFA201985	E1M10000236E02	ECO102340 ECO103878	S1M10000006C06	
E3M10000004D01	EFA201983	E1M10000236E02	ECO103878 ECO204942	S1M10000006D06	SAU802496
			ECO204942 ECO103287	S1M10000006F06	SAU802490
E3M10000004D01	EFA202953	E1M10000236A03	ECO103287 ECO102556	S1M10000006F06	SAU800006
E3M10000004G01	EFA200839	E1M10000236D03 E1M10000236G03	ECO102556 ECO102655	S1M10000006G08	SAU800006 SAU800967
E3M10000004D02	EFA202022		ECO102655 ECO103186	S1M10000006A07	SAU800967 SAU801760
E3M10000004D02	EFA202028	E1M10000236A04	ECO103180	311411000000000007	BAU001700

E3M10000004C03 EFA200412 E E3M10000004A04 EFA201981 E E3M10000004A04 EFA205229 E E3M10000004F08 EFA201977 E E3M10000004F08 EFA203137 E	Clone Name E1M10000236D04 E1M10000236G04 E1M10000236F05 E1M10000236F05 E1M10000236H06 E1M10000236H06 E1M10000236D08 E1M10000236F09 E1M10000236C10	Gene LocusID ECO103481 ECO103510 ECO102847 ECO103181 ECO103182 ECO103242 ECO103243 ECO103669	Clone Name S1M10000006C07 S1M10000006D07 S1M10000006G07 S1M10000006A08 S1M10000006E08 S1M10000006A10	Gene LocusID SAU800546 SAU801105 SAU802496 SAU802496 SAU802238 SAU802496
E3M10000004D02 EFA202536 E E3M10000004C03 EFA200412 E E3M10000004A04 EFA201981 E E3M10000004A04 EFA205229 E E3M10000004F08 EFA201977 E E3M10000004F08 EFA203137 E	E1M10000236G04 E1M10000236A05 E1M10000236F05 E1M10000236F05 E1M10000236H06 E1M10000236H06 E1M10000236D08 E1M10000236F09 E1M10000236C10	ECO103481 ECO103510 ECO102847 ECO103181 ECO103182 ECO103242 ECO103243 ECO103669	\$1M1000006D07 \$1M1000006E07 \$1M10000006G07 \$1M10000006A08 \$1M10000006E08 \$1M10000006A10	SAU800546 SAU801105 SAU802496 SAU801731 SAU802496 SAU802238
E3M10000004C03 EFA200412 E E3M10000004A04 EFA201981 E E3M10000004A04 EFA205229 E E3M10000004F08 EFA201977 E E3M10000004F08 EFA203137 E	E1M10000236G04 E1M10000236A05 E1M10000236F05 E1M10000236F05 E1M10000236H06 E1M10000236H06 E1M10000236D08 E1M10000236F09 E1M10000236C10	ECO103510 ECO102847 ECO103181 ECO103182 ECO103242 ECO103243 ECO103669	\$1M1000006D07 \$1M1000006E07 \$1M10000006G07 \$1M10000006A08 \$1M10000006E08 \$1M10000006A10	SAU801105 SAU802496 SAU801731 SAU802496 SAU802238
E3M10000004A04 EFA201981 E E3M10000004A04 EFA205229 E E3M10000004F08 EFA201977 E E3M10000004F08 EFA203137 E	E1M10000236A05 E1M10000236F05 E1M10000236F05 E1M10000236H06 E1M10000236H06 E1M10000236D08 E1M10000236F09 E1M10000236C10	ECO102847 ECO103181 ECO103182 ECO103242 ECO103243 ECO103669	S1M10000006E07 S1M10000006G07 S1M10000006A08 S1M10000006E08 S1M10000006A10	SAU802496 SAU801731 SAU802496 SAU802238
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E3M10000004F08 EFA203137 E	E1M10000236H06 E1M10000236H06 E1M10000236D08 E1M10000236F09 E1M10000236C10	ECO103242 ECO103243 ECO103669	S1M10000006E08 S1M10000006A10	SAU802238
251110000001100	E1M1000236H06 E1M10000236D08 E1M10000236F09 E1M10000236C10	ECO103243 ECO103669	S1M10000006A10	
- 1 100 X (1 0 0 0 0 0 0 0 0 1 0 1 0 1 0 1 0 1 0	E1M10000236D08 E1M10000236F09 E1M10000236C10	ECO103669		・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・・
131111000000 1210	E1M10000236F09 E1M10000236C10		S1M10000006B10	SAU802240
1,51411000000 12510	E1M10000236C10		S1M10000000B10	SAU802496
D31/11/00/00/00/11 10 ====================		ECO103228	S1M10000006G10	SAU802247
ESIMIOGOGO ESTA		ECO102227	S1M10000000G10	SAU802248
1331111000000 12111	E1M10000236A11	ECO102986	S1M10000006B11	SAU801618
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Belitious	E1M10000237B02	ECO101830	S1M10000006A12	SAU800348
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1 1111111111111111111111111111111111111	E1M10000237E04	ECO103218	S1M10000007F01	SAU801256
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	E1M10000237A08	ECO103216	S1M10000007E04	SAU802496
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E3M10000005C04 EFA200661	E1M10000237B08	ECO101186	S1M10000007C05	SAU800547
	E1M10000237D08	ECO103217	S1M10000007G05	SAU800548
	E1M10000237D08	ECO103216	S1M10000007C06	SAU801900
E3M10000005G05 EFA201977	E1M10000237E08	ECO103262	S1M10000007D06	SAU800547
E3M10000005G05 EFA203137	E1M10000237E08	ECO103878	S1M10000007E06	SAU801113
E3M10000005A07 EFA200811	E1M10000237E08	ECO204942	S1M10000007C07	SAU801904
E3M1000005A07 EFA200812	E1M10000237B09	ECO101844	S1M10000007E07	SAU801618
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E3M10000005B08 EFA201977	E1M10000237D10		S1M10000007C08	SAU800482
E3M10000005B08 EFA203137	E1M10000237E11		S1M10000007E08	SAU800700
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E3M10000005D10 EFA201977	E1M10000238A02		<u> </u>	SAU800210
E3M10000005D10 EFA203137	E1M10000238F03		S1M10000007D10	SAU800537
E3M10000005F10 EFA201977	E1M10000238B04		S1M10000007F10	· SAU802240
E3M10000005F10 EFA203137	E1M10000238B04		S1M10000007B11	SAU802177
E3M10000005110 EFA201982	E1M10000238B04		S1M10000007B11	SAU802176
E3M1000000C01 EFA201981	E1M10000238D04		S1M10000007D11	SAU801900
E3M1000000G02 EFA202214	E1M10000238F04			SAU802160
E3M1000000G02 EFA202214 E3M1000006G02 EFA202216	E1M10000238F04		1'	SAU603460
E3M10000006B03 EFA202210	E1M10000238F04			SAU800519
	E1M10000238E03		<u> </u>	SAU802643
	E1M10000238F05		S1M10000008A03	SAU802177
	E1M10000238F03			SAU802176
				SAU800023
E3M10000006F04 EFA200811	E1M10000238D06			SAU800753
E3M1000006F04 EFA200812	E1M10000238F06		.(SAU802369
E3M1000006G04 EFA201999	E1M10000238F06	ECO103230	311411000000000000	DAC602309

Class Name	Gana	Clone Name	Gene	Clone Name	Gene
Clone Name	Gene LocusID	Cione Ivanie	LocusID		LocusID
E3M10000006G04	EFA201997	E1M10000238A07	ECO103236	S1M10000008A04	SAU800478
E3M10000006H09	EFA201028	E1M10000238A07	ECO103237	S1M10000008B04	SAU802496
E3M10000006E11	EFA200811	E1M10000238A08	ECO101628	S1M10000008D05	SAU800517
E3M10000006E11	EFA200812	E1M10000238E08	ECO103237	S1M10000008D05	SAU202623
E3M10000006C12	EFA205225	E1M10000238E08	ECO103238	S1M10000008E05	SAU801183
E3M10000006C12	EFA201977	E1M10000238B09	ECO102213	S1M10000008G05	SAU800305
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E3M10000033B09						SAU802194
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E3M10000033D99 EFA200621 E1M10000268008 EC0103223 SIM10000018G08 SAU802586 E3M10000033G09 EFA200457 E1M100000268008 EC0103243 SIM10000018G08 SAU802587 E3M10000033G09 EFA201028 E1M10000268008 EC0103243 SIM10000018G08 SAU802587 E3M10000033H09 EFA201028 E1M10000268B08 EC0103243 SIM10000018G08 SAU802587 E3M10000033H09 EFA201028 E1M10000268B08 EC0103263 SIM10000018A09 SAU800217 E3M10000033C10 EFA201986 E1M10000268B09 EC0103263 SIM10000018B09 SAU801843 E3M10000033D10 EFA203255 E1M10000268B09 EC0103263 SIM10000018B09 SAU801843 E3M10000033D10 EFA203256 E1M10000268B09 EC0103263 SIM10000018B09 SAU801843 E3M10000033D10 EFA20326 E1M10000268F09 EC0103223 SIM10000018C09 SAU801843 E3M10000033D10 EFA202012 E1M10000268F09 EC0103223 SIM10000018C09 SAU801753 E3M10000033D11 EFA20217 E1M10000268F09 EC0103227 SIM10000018D09 SAU801751 E3M10000033D11 EFA20217 E1M10000268F09 EC0103227 SIM10000018D09 SAU802554 E3M10000033D11 EFA202017 E1M10000268F09 EC0103237 SIM10000018D09 SAU802554 E3M10000033D11 EFA202017 E1M10000268F09 EC0103237 SIM10000018D09 SAU802555 E3M10000033D11 EFA202013 E1M10000268B10 EC0103238 SIM10000018D09 SAU802555 E3M10000033D11 EFA202013 E1M10000268B10 EC0103238 SIM10000018G09 SAU802555 E3M10000033D11 EFA202013 E1M10000268B10 EC0103233 SIM10000018G09 SAU802555 E3M10000033D11 EFA202031 E1M10000268B11 EC0101812 SIM10000018G09 SAU802555 E3M10000033D12 EFA200812 E1M10000268B11 EC0103232 SIM10000018G09 SAU802555 E3M10000033D12 EFA200812 E1M10000269D01 EC0103231 SIM1000018G09 SAU802555 E3M10000033D12 EFA200812 E1M10000269D01 EC0103232 SIM1000018G09 SAU802555 E3M10000033D12 EFA200815 E1M10000269D01 EC0103232 SIM10000018D10 SAU802555 E3M10000033D12 EFA200815 E1M10000269D01 EC0103232 SIM10000018D10 SAU802555 E3M10000034D02 EFA202036 E1M10000269D04 EC0103231 SIM1000018D10 SAU802554 E3M10000034D02 EFA202036 E1M10000269D04 EC0103231 SIM10000018D10 SAU802654 E3M10000034D02 EFA202036 E1M10000269D04 EC0103231 SIM10000018D10 SAU802654 E3M10000034D04 EFA202030 E1M10000269D05 EC0102323 SIM10000018D11 SAU802654 E3M10000034D04 EFA202030 E						SAU802654
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B3M10000033C11						
E3M10000033B11						
E3M10000033B11	1					
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Clone Name	Gene LocusID	Clotte Ivaille	LocusID	Cione ivanic	LocusID
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E3M10000052C01	EFA202110			1	SAU800517
E3M10000052F01	EFA200457			1	SAU202623
E3M10000052C02	EFA200805				SAU202023 SAU801790
E3M10000052D02	EFA202170				
E3M10000052G02	EFA200457				SAU800529
E3M10000052B03	EFA200239			1	SAU801790
E3M10000052E03	EFA202378				SAU801139
E3M10000052G03	EFA200326		l .	·	SAU800266
E3M10000052B04	EFA200290	E1M10000291G05	ECO102556	S1M10000029H10	SAU801139

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	EFA200198	E1M10000291C00	EC0100102	S1M10000029A12	SAU801631
E3M10000052C05	EFA201888	E1M10000291E07	ECO103866	S1M10000029C12	SAU802070
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E3M10000032F03	EFA202108	E1M10000291B08	EC0103217	S1M10000029F12	SAU800253
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E3M10000052B07	EFA202108 EFA202378	E1M10000291E10	ECO101591	S1M10000030D01	SAU801113
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	EFA200240 EFA200898	E1M10000291H11	ECO103202	S1M10000030C02	SAU801515
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	ECO100023 ECO100702	E1M10000291B12	ECO103882 ECO103243	S1M10000030H02	SAU802654
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1053-37	ECO101238	E1M10000293B01	ECO103883	S1M10000030C03	SAU801473
1053-37		E1M10000293B02	ECO104093	S1M10000030B03	SAU800542
1010-C11	ECO101324	E1M10000293G02	ECO103880 ECO100402	S1M10000030H03	SAU800232
1017-H1	ECO304472	E1M10000293A04 E1M10000293B04	ECO100402 ECO103886	S1M10000030103	SAU800232 SAU800526
1067-16	ECO102309	E1M10000293B04 E1M10000293A05	ECO103880 ECO100095	S1M10000030C04	,SAU800478
1083-27	ECO102636 ECO102557	E1M10000293A03 E1M10000293E05	ECO100093 ECO103223	S1M10000030A03	SAU801256
1065-12	ECO102337	E1M10000293E05	ECO103224	S1M10000030D03	SAU800526
221-41	ECO103884	E1M10000293E03	ECO103224 ECO103243	S1M10000030C03	SAU800759
B17-6.O10		E1M10000293G03	ECO103243	S1M10000030D05	SAU302793
910-B20	ECO103884	E1M10000293H06	ECO101173	S1M1000030D05	SAU800776
B18-2.N21	ECO100148	E1M10000293F07	ECO102034 ECO101095	S1M10000030G05	SAU800777
971-B20	ECO103240	E1M10000293F07	ECO101093 ECO101844	S1M10000030H05	SAU800179
971-B20	ECO103241			S1M10000030H03	SAU800179 SAU800189
D1-1.A15	ECO103394	E1M10000293E08	ECO101939 ECO103101	S1M10000030D06	SAU801257
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PJMF55	ECO103265	E1M10000293A11	ECO103883	S1M10000030C08	SAU801831 SAU802231
R1-15.A13	ECO101995	E1M10000293E11	ECO103242	S1M10000030F08	
R1-19.H1	ECO101104	E1M10000293F11	ECO104091	S1M10000030F08	SAU802230
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709-F23	ECO101506	E1M10000295G01	ECO103532	S1M10000030F09	SAU801904
801-C15	ECO100488	E1M10000295G01	ECO103533	S1M10000030G09	SAU800542
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801-C15	ECO100491	E1M10000295E02	ECO103217	S1M10000030A10	SAU802309
801-H19	ECO100488	E1M10000295E02	ECO103218	S1M10000030A10	SAU802308
801-H19	ECO100490	E1M10000295F02	ECO100169	S1M10000030C10	SAU800562

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Cione Name	LocusID	Cione Ivallie	LocusID	Clone I value	LocusID
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804-P6	ECO102513	E1M10000295A07	ECO100712	S1M10000030F10	SAU800019
807-D20	ECO100366	E1M10000295B07	ECO100179	S1M10000030G10	SAU800019
807-D20	ECO100367	E1M10000295B07	ECO100180	S1M10000030H10	SAU802654
B13-17.G8	ECO100307	E1M10000295C07	ECO103224	S1M10000030A11	SAU800517
	ECO101111	E1M10000295C07	ECO103225	S1M10000030A11	SAU202623
B5-6.C8 B5-6.C8	ECO101475	E1M10000295C07	ECO103226	S1M10000030D11	SAU800517
ii	ECO201962	E1M10000295C07	ECO103225	S1M10000030D11	SAU202623
B5-6.C8		E1M10000295D08	ECO103225	S1M10000030E11	SAU802241
B8-2.D9	ECO103461	E1M10000295F08	ECO103220	S1M10000030E11	SAU800811
B15-8.P13	ECO101328	E1M10000295F08	ECO103100 ECO103217	S1M10000030G11	SAU801647
B15-8.P13	ECO101329		ECO103217	S1M10000030C12	SAU801646
T13-5.A2	ECO103059	E1M10000295G08	ECO103218	S1M10000030E12	SAU800537
T12-3.I11	ECO102857	E1M10000295B09		S1M10000030E12	SAU800557
T20-15.D4	ECO101475	E1M10000295F09	ECO103881	S1M10000030G12	SAU801320
T20-15.D4	ECO101476	E1M10000295F09 E1M10000295G09	ECO103882 ECO103263	S1M10000031B01	SAU800023
T20-15.D4	ECO201962			S1M10000031H01	SAU800023
T24-15.G6	ECO103059	E1M10000295D10	ECO103101	S1M10000031B02	'SAU801912
T24-17.C6	ECO102857	E1M10000295H10	ECO103263	S1M10000031E02	SAU801912 SAU802231
244.B12	ECO101763	E1M10000295B11	ECO103229	S1M10000031F02	SAU802231
244.B12	ECO101764	E1M10000295F11	ECO100954 ECO103494	S1M10000031F02	SAU802235
244.B12	ECO101765	E1M10000295G12			SAU802233 SAU802234
1042-J1	ECO100702	E1M10000312D11	ECO104091	S1M10000031G02	SAU801355
1042-J1	ECO100703	E1M10000312D11	ECO104092	S1M10000031H02	
195.F5	ECO102842	E1M10000296B01	ECO102304	S1M10000031A03	SAU802250
25.D5	ECO103059	E1M10000296C02	ECO102466	S1M10000031E03	SAU801134
25.D6	ECO103059	E1M10000296C02	ECO102467	S1M10000031E03	SAU801135
177.F3	ECO102309	E1M10000296D02	ECO103235	S1M10000031F03	SAU802240
525.H11	ECO102857	E1M10000296D02	ECO103236	S1M10000031G03	SAU801505
632.N2	ECO104277	E1M10000296D02	ECO103237	S1M10000031A04	SAU801434
633.B7	ECO103479	E1M10000296H02	ECO102556	S1M10000031A04	SAU302892
671.I20	ECO103478	E1M10000296C03	ECO100150	S1M10000031B04	SAU800543
676.B12	ECO103479	E1M10000296C03	ECO100151	S1M10000031C04	SAU800738
643.B19	ECO100702	E1M10000296E03	ECO101086	S1M10000031C04	SAU800737
720.O16	ECO103884	E1M10000296H03	ECO103227	S1M10000031E04	SAU800542
666.H12	ECO103478	E1M10000296H03			SAU801517
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844.B21	ECO102144	E1M10000296F05	1	S1M10000031G04	SAU302882
P31-25-F3	ECO101686	E1M10000296G05	·	S1M10000031F05	SAU800548
P335-8.H8	ECO101041	E1M10000296H05			SAU801526
P347.2	ECO101086	E1M10000296A06		S1M10000031G06	SAU800548
P31-11-J20	ECO103228	E1M10000296A06			SAU600582
P336-14.F20	ECO101370	E1M10000296G07		<u> </u>	SAU801760
P31-27-M1	ECO103423	E1M10000296G07	l		SAU801181
P338-4.M21	ECO100139	E1M10000296H07	ECO103220		SAU800016
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P31-2-E16	ECO101686	E1M10000296E08	<u> </u>		SAU801790
P335-3.J14	ECO100523	E1M10000296F08	I	S1M10000031E08	SAU800547
P334-5.H2	ECO100139	E1M10000296G08			SAU801264
P31-33-N2	ECO103241	E1M10000296H08			SAU801193
P332-11.C20	ECO102827	E1M10000296H08		S1M10000031D09	SAU800019
P332-11.C20	ECO102828	E1M10000296A09			SAU800006
869.A23	ECO100390	E1M10000296B11	ECO103229	S1M10000031H09	SAU801599

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P326-9.K2		E1M10000296E11	ECO101684	S1M10000031E10	SAU800001
P326-9.K2	ECO103293	E1M10000290F12	ECO101084	S1M10000031E10	SAU800244
P323-8.P1	ECO101685	E1M10000298C01	ECO100033	S1M10000031G10	SAU800962
P35-8	ECO103692	E1M10000298C01	ECO101438	S1M10000031G10	SAU801741
P36-13.E2	ECO103059	E1M10000298G01	ECO104148	S1M10000031A11	SAU801908
P38-1.G20	ECO102227		ECO104149	S1M10000031B11	SAU802152
P327-50.M10	ECO103242	E1M10000298G02	ECO102030	S1M10000031E11	SAU800312
P327-50.M10	ECO103243	E1M10000298C03		S1M10000031F11	SAU801234
P328-8.D21	ECO103240	E1M10000298C03	ECO103239	S1M10000031H11	SAU800962
P328-8.D21	ECO103241	E1M10000298D03	ECO103886	S1M10000031H11	SAU801621
P328-20.P20	ECO100541	E1M10000298H03	ECO103262	S1M10000031B12	SAU801021
P33-1.C22	ECO103227	E1M10000298H03	ECO103878	S1M10000031E12	SAU801741
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X3S107-17	ECO101476	E1M10000298E04	ECO100430	S1M10000031F12	SAU800244 SAU802654
X3S107-17	ECO201962	E1M10000298E04	ECO100431		SAU802034 SAU800548
P35-7	ECO103928	E1M10000298H04	ECO100809	S1M10000032C01	
X3S118-9	ECO103263	E1M10000298H04	ECO100808	S1M10000032F01	SAU800525
X3S163-1	ECO103423	E1M10000298C05	ECO103234	S1M10000032F01	SAU800524
X3S204-7	ECO103238	E1M10000298C05	ECO103235	S1M10000032H01	SAU802112
X3S177-4	ECO101161	E1M10000298C05	ECO103236	S1M10000032H01	SAU802111
P342-3	ECO102104	E1M10000298D05	ECO101539	S1M10000032E02	SAU801096
SC21.1	ECO103224	E1M10000298D05	ECO101540	S1M10000032G02	SAU800830
SC17.1	ECO102087	E1M10000298C06	ECO101844	S1M10000032G02	SAU800829
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SC13.1	ECO101348	E1M10000298G06	ECO100096	S1M10000032C03	SAU800771
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E1M10000197H10	ECO100974	P1M10000073G03	l	S1M10000040F04	SAU800525
E1M10000197A11	ECO100974	P1M10000073D04	L	S1M10000040G04	SAU800517
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Clone Name	Gene LocusID	Cione Ivallie	LocusID	Clone I value	LocusID
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E1M10000198C10	ECO103912	P1M10000076D05	PAE201875	S1M10000040H07	SAU802082
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E1M10000201G01	ECO103243	P1M10000082A02	PAE202946	S1M10000041H05	SAU801246
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	ECO103234 ECO103230	P1M10000104A02	PAE200887	S1M10000043B02	SAU802262
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E11V110000208H07	ECO1038/8	F11V11UUUU1U3DU4	FAEZ00303	511/11/00/0435/05	SAUGUITAL

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				PAE200221	J	
E1M10000213F07 ECO104168 P1M10000099F10 PAE200423 S1M10000044D04 SAU802238			<u> </u>			
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E1M10000229G05	E1M10000229E03	ECO103237		L		
E1M10000229H06		ECO103238			I *	
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1 11111000002331103		l			

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Table IC provides a cross reference between PathoSeq Gene Loci listed in Table IB and the SEQ ID NOs. of the corresponding PathoSeq polypeptides and the SEQ ID NOs; of the nucleic acids which encode them. The Gene Locus IDs provided in Table IC each comprise a nine digit alpha-numeric identifier that can be used to determine the organism from which each Gene Locus and corresponding SEQ ID NOs. were identified. Specifically, the first letter of the Gene Locus ID corresponds to the first letter of the genus name of the organism described herein from which the Gene Locus was identified and the second and third letters of the Gene Locus ID correspond to the first two letters of the species name of this organism. For example, the identifier EFA205257 describes a gene locus identified from *Enterococcus faecalis*. In those instances where the three letter identifier is the same for different organisms, the exact identity of the organism which corresponds to the Gene Locus ID can be determined by referring to the organism designation in the sequence listing for the coding nucleic acid or polypeptide SEQ ID NO. that corresponds to the particular Gene Locus ID.

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TABLE IC

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6217	42401	EFA201977	18279	54463	CJU100856	30340	66524	PAE203658
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6230	42414	EFA201975	18292	54476	CJU100890	30353	66537	PAE203741
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6233	42417	EFA201985	18295	54479	СЛU100903	30356	66540	PAE203744
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6236	42420	EFA202022	18298	54482	CJU100929	30359	66543	PAE203755
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6261	42445	EFA202012	18323	54507	CJU101022	30384	66568	PAE203892
6262	42446	EFA202007	18324	54508 54500	CJU101027	30385	66569	PAE203897
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6264 6265		EFA201888	18326	54510 54511	CJU101029	30387	66571	PAE203900
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6324	42508	EFA202162 EFA204646	18386	54570	CJU101217	30447	66631	PAE204281
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6332	42516	EFA201041	18394	54578	CJU101290	30455	66639	PAE204357
6333	42517	EFA201801	18395	54579	СЈU101295	30456	66640	PAE204359
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6689 42873 ECO101780 18751 54935 CPN200879 30812 66996 PMU100435 6690 42874 ECO101637 18752 54936 CPN200887 30813 66997 PMU100438 6691 42875 ECO101406 18753 54937 CPN200889 30814 66998 PMU100439 6692 42876 ECO100153 18754 54938 CPN200890 30815 66999 PMU100440 6693 42877 ECO100113 18755 54939 CPN200891 30816 67000 PMU100444 6694 42878 ECO101700 18756 54940 CPN200893 30817 67001 PMU100448 6695 42879 ECO103160 18758 54942 CPN200895 30818 67002 PMU100453 6696 42880 ECO103161 18759 54943 CPN200895 30820 67004 PMU100456 6698 42882 ECO10364 18760 54944 CPN200903<	6687	42871	ECO101679	18749	54933	CPN200876	30810	66994	PMU100425
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6766	42950	ECO102175	18828	55012	CTR100036	30889	67073	PMU100719
6767	42951	ECO102554	18829	55013	CTR100067	30890	67074	PMU100720
6768	42952	ECO100169	18830	55014	CTR100083	30891	67075	PMU100722
6769	42953	ECO102158	18831	55015	CTR100183	30892	67076	PMU100726
6770	42954	ECO103886	18832	55016	CTR100201	30893	67077	PMU100729
6771	42955	ECO103466	18833	55017	CTR100305	30894	67078	PMU100730
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6775	42959	ECO103202 ECO100201	18837	55020	CTR100407 CTR100501	30897	67081	PMU100730
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6779	42963	ECO202164	18841	55025	CTR100660	30902	67086	PMU100755
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6810	42994	ECO103881	18872	55056	CTR200064	30933	67117	PMU100869
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6812	42996	ECO102274	18874	55058	CTR200070	30935	67119	PMU100873
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6814	42998	ECO101524	18876	55060	CTR200076	30937	67121	PMU100875
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6819	43003	ECO103366	18881	55065	CTR200088	30942	67126	PMU100893
6820	43004	ECO102294	18882	55066	CTR200096	30943	67127	PMU100894
6821	43005	ECO102295	18883	55067	CTR200100	30944	67128	PMU100897
6822	43006	ECO101669	18884	55068	CTR200102	30945	67129	PMU100898
6823	43007	ECO103331	18885	55069	CTR200103	30946	67130	PMU100903
6824	43008	ECO103725	18886	55070	CTR200104	30947	67131	PMU100904
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6826	43010	ECO100757	18888	55072	CTR200112	30949	67133	PMU100917
6827	43011	ECO101139	18889	55073	CTR200114	30950	67134	PMU100920
6828	43012	ECO104048	18890	55074	CTR200116	30951	67135	PMU100924
6829	43013	ECO100468	18891	55075	CTR200123	30952	67136	PMU100936
6830	43014	ECO100659	18892	55076	CTR200124	30953	67137	PMU100938
6831	43015	ECO100456	18893	55077	CTR200126	30954	67138	PMU100941
6832	43016	ECO103483	18894	55078	CTR200127	30955	67139	PMU100945
6833	43017	ECO103536	18895	55079	CTR200130	30956	67140	PMU100948
6834	43018	ECO100555	18896	55080	CTR200132	30957	67141	PMU100949
6835	43019	ECO103756	18897	55081	CTR200133	30958	67142	PMU100950
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8525	44709	ABA100914	20587	56771	ECO103822	32648	68832	PRT102791
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8527	44711	ABA100920	20589	56773	ECO103852	32650	68834	PRT102802
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8630	44814	ABA101608	20692	56876	EFA200155	32753	68937	PRT103781

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8903 8904	45088	ABA104774 ABA104775	20965	571 <b>49</b> 571 <b>5</b> 0	EFA201465 EFA201466	33026 33027	69210	PRT106097 PRT106098
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8970	45154	ABA105208 ABA105243	21032 21033	57216 57217	EFA201812 EFA201824	33093 33094	69277 69278	PSY100759 PSY100792
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9074	45258	ABA106040	21136	57320	EFA202357	33197	69381	PSY101698
9075	45259	ABA106047	21137	57321	EFA202358	33198	69382	PSY101701
9076	45260	ABA106057	21138	57322	EFA202359	33199	69383	PSY101711
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9078	45262	ABA106059	21140	57324	EFA202362	33201	69385	PSY101745
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10901   47085   BBU100302   22963   59147   KPN205236   35024   71208   SEP202218   10902   47086   BBU100303   22964   59148   KPN205380   35025   71209   SEP202219   10904   47088   BBU100310   22966   59150   KPN205542   35027   71211   SEP202225   10905   47089   BBU100313   22967   59151   KPN205579   35028   71212   SEP202228   10906   47090   BBU100312   22968   59152   KPN205610   35029   71213   SEP202229   10907   47091   BBU100325   22969   59153   KPN206188   35030   71214   SEP202223   10909   47092   BBU100333   22970   59154   KPN206317   35032   71216   SEP202232   10909   47093   BBU100334   22971   59155   KPN206317   35032   71216   SEP202232   10909   47094   BBU100336   22972   59156   KPN206346   35033   71217   SEP202229   10912   47096   BBU100337   22973   59157   KPN206682   35034   71218   SEP202229   10912   47096   BBU100337   22973   59157   KPN206682   35034   71218   SEP202239   10912   47098   BBU100337   22975   59159   KPN2066674   35036   71220   SEP202312   10913   47097   BBU100340   22976   59160   KPN206801   35037   71221   SEP202387   10915   47099   BBU100341   22977   59161   KPN207037   35038   71223   SEP202395   10916   47100   BBU100344   22978   59162   KPN207078   35039   71223   SEP202340   10916   47100   BBU100346   22998   59164   KPN207172   35041   71225   SEP202340   10920   47104   BBU100367   22988   59166   KPN207129   35041   71225   SEP202340   10920   47104   BBU100367   22988   59166   KPN207129   35044   71226   SEP202354   10922   47106   BBU100367   22988   59168   KPN207142   35040   71224   SEP202351   10924   47105   BBU100369   22985   59166   KPN207142   35041   71225   SEP202354   10924   47105   BBU100367   22988   59168   KPN207142   35041   71225   SEP202354   10924   47108   BBU100367   22988   59168   KPN207142   35041   71225   SEP202550   10924   47106   BBU100367   22988   59168   KPN207142   35044   71226   SEP202550   10924   47108   BBU100374   22980   59175   KPN300043   35057   71234   SEP202555   10925   47109   BBU100388									
10902									
10903						1			
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10907									
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